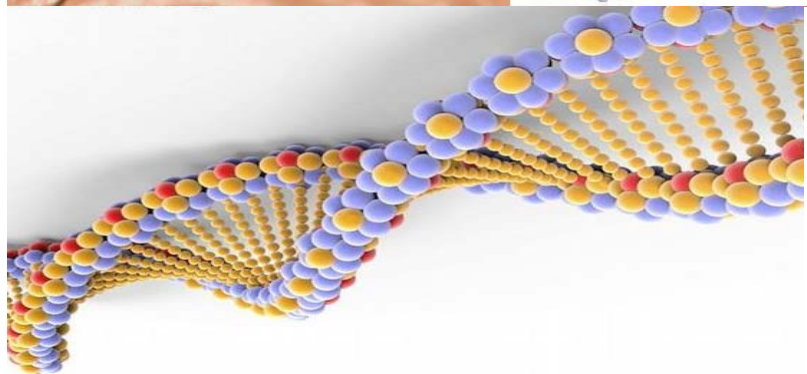
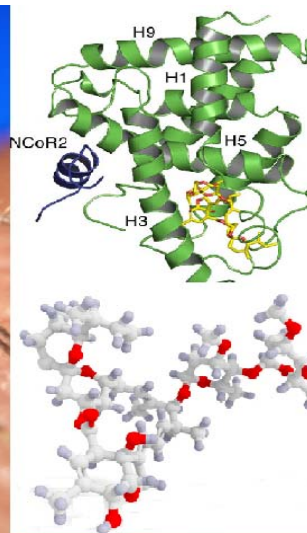


PhytoChem & BioSub Journal

Peer-reviewed research journal on Phytochemistry & Bioactives Substances

ISSN 2170 - 1768



PCBS Journal

Volume 10 N° 3

2016



PhytoChem & BioSub Journal

ISSN 2170 – 1768

Peer-reviewed research journal on Phytochemistry & Bioactives Substances

CAS Source Index (CODEN: PBJHB3)

Editor in Chief

Pr Abdelkrim CHERITI

Phytochemistry & Organic Synthesis Laboratory
08000, Bechar, Algeria

PhytoChem & BioSub Journal (PCBS Journal) is a peer-reviewed research journal published by Phytochemistry & Organic Synthesis Laboratory. The PCBS Journal publishes innovative research papers, reviews, mini-reviews, short communications and technical notes that contribute significantly to further the scientific knowledge related to the field of Phytochemistry & Bioactives Substances (Medicinal Plants, Ethnopharmacology, Pharmacognosy, Phytochemistry, Natural products, Analytical Chemistry, Organic Synthesis, Medicinal Chemistry, Pharmaceutical Chemistry, Biochemistry, Computational Chemistry, Molecular Drug Design, Pharmaceutical Analysis, Pharmacy Practice, Quality Assurance, Microbiology, Bioactivity and Biotechnology of Pharmaceutical Interest). Contributions in all areas at the interface of Chemistry, Pharmacy, Medicine and Biology are welcomed.

Submission of an article to the *PCBS Journal* implies that the work described has not been published previously (except in the form of an abstract or as part of a published lecture or academic thesis), that it is not under consideration for publication elsewhere, that its publication is approved by all authors.

The *PCBS Journal* reserves the right to submit all received manuscripts to *ad hoc* referees, whose names will be kept confidential, and will have the authority to decide on the pertinence for acceptance. Referees may send back manuscripts to Editor-in-Chief, for transmission to the author(s) with suggestions for necessary alterations, which are to be made in order to conform to the standards and editorial rules of the Journal. All manuscripts should be prepared in MS-Word format, and submitted online to **Phytochem07@yahoo.fr**. Upon receipt of paper submission, the Editor sends an E-mail of confirmation to the corresponding author within 1-4 working days. The Editors reserve the right to edit or otherwise alter all contributions, but authors will receive proofs for approval before publication.

Editorial Board

Abou Enein H

Pharm. Med Chem Dept. Research Division,
NRC, Dokki, Giza, Egypt

Allali H.

LASNABIO, Dept. Chemistry,
University of Tlemcen, Algeria

Awad Allah A.

Dept. Chem., Faculty of Science, Islamic
University of Gaza, Gaza, Palestine

Barkani M.

Materials Laboratory, Bedjai University,
Algeria

Benharathe N

Materials Laboratory, USTO university, Oran,
Algeria

Boksha I.

Federal Research Centre for Epidemiology
Microbio., MH, Moscow, Russia

Boukir A.

Lab. Applied Chem., Faculty of Science,
S.M.Ben Abdellah Univ., Fez, Morocco

Boulenouar N.

Biochemical Laboratory, Nour E. University, El
Bayadh, Algeria

Daoud K.

GP- Indus.Pharma Laboratory, USTHB, Algiers,
Algeria

El Abed D.

Fine Organic Chemistry laboratory, Es Senia
university, Oran, Algeria

El Omar F.

Applied Chem. Lab., Faculty of Science
Lebanese University, Tripoli, Lebanon

Govender P.

KwaZulu-Natal Univ., School of Life Sci.
Biochem., Durban, South Africa

Gargouri A. F.

Biotechnology center, CBS
Sfax, Tunisia

Gherraf N.

LRNAMS Laboratory, Larbi ben M'hidi,
University, Oum El-Bouaghi, Algeria

Gouasmia A.

Organic Materials Laboratory, faculty of science,
Tebessa University, Algeria

Kajima J.M

COSNA Laboratory, faculty of science, Tlemcen
University, Algeria

Khelil-Oueld Hadj A.

ECOSYS Laboratory, Ouargla, University,
Ouargla, Algeria

Marouf A.

Biochemistry laboratory, Dept of Biology,
Naama University, Algeria

Laouar H

NRV laboratory, Dept. Biology and plant
ecology, F.A. University, Setif-I, Algeria

Oueld Hadj M.D.

ECOSYS Laboratory, Ouargla, University,
Ouargla, Algeria

Roussel C.

Chirosciences, UMR 7313, Stereo-Dyna.
Chiralty, Aix-Marseille Univ., France

Sidiqi S. K.

Bioorganometallic Lab., Dept. chemistry, AMU
University, New Delhi, India

Tabti B.

LASNABIO, Dept. Chemistry, University of
Tlemcen, Algeria

Youcefi M.

LSF laboratory, faculty of sciences, Laghouat
University, Algeria

Afaxantidis J.

Synerlab Développement,
Orléans, France

Allouch A.

Applied Chem. Lab., Faculty of Science Lebanese
University, Tripoli, Lebanon

Badjah A.Y.

Dept. Chem., College of Science,
King Saud Univ., Riyadh, KSA

Belboukhari N.

LMBSC Lab. Bechar university
Algeria

Bennaceur M.

Biochemical Laboratory, Biology faculty, Es Senia
University, Oran, Algeria

Bouchekara M.

Chemistry Laboratory, Science faculty, University of
Mascara, Algeria

Brada M.

Valuation of Natural Substances Lab., Khemis-
Miliana University, Algeria

Dadamoussa B.

Chemistry Laboratory, Ghardai University,
Algeria

Djebar S.

Materials & mineral laboratory, USTHB, Algiers,
Algeria.

Elachouri M.

Lab. Physiology and Ethnopharma...Sci.Fac Med. I
University. Oujda, Morocco

Ermel G.

Rennes University EA 1254, Beaulieu Campus
Rennes, France

Hacini S.

Fine Organic Chemistry laboratory, Es Senia
university, Oran, Algeria

Ghanmi M.

Medicinal plants division, CRF, Agdal
, Rabat, Morocco

Ghezali S.

IAP, Dept Catalysis, Sonatrach, Algiers,
Algeria

Kabouche Z.

LOST Laboratory, faculty of sciences, Constantine
University, Algeria

Kaid-Harche M.

Biotechnology Laboratory, Faculty of biology,
USTO, Oran, Algeria

Lahreche M.B.

LCO laboratory, faculty of Biology, Djelfa
University, Algeria

Meddah B.

Lab. Pharmaco. Toxic. Faculty of medicine and
pharmacy, Med. V Univ. Rabat, Morocco

Mushfik M.

Natural products laboratory, Dept chemistry, AMU
university, New Delhi, India

Rahmouni A.

LMC laboratory, Dept Chemistry, Saida University,
Algeria

Saidi M.

LPSEZA laboratory, Dept Chemistry, Ouargla
University, Algeria

Soltani Y.

BPO Laboratory, Endocrinology team, Dept. Bio.
Physio., USTHB, Algiers, Algeria,

Taleb S.

Materials Chemistry Laboratory
Dept Chem. UDL Univ., SBA, Algeria

Akkal S.

Research Unity: VNRBM Lab. Dept. Chem.,
University of Constantine 1, Algeria

Aouf N.

Lab. Applied Org. Chem. , Dpt. Chem.,
Annaba University, Algeria

Balansard G.

Pharmacognosy Lab., Faculty of pharmacy, Univ.
Aix Marseille II, Marseille, France

Belkhiri A.

Pharmacognosy Laboratory, Faculty of Medicine,
Constantine university, Algeria

Berredjem M.

Lab. Applied Org. Chem. , Dpt. Chem.,
Annaba University, Algeria

Bouklouze A.

Lab. Pharmaco. Toxic. Faculty of medicine and
pharmacy, Med. V Univ. Rabat, Morocco

Bressy C.

iSm2, CNRS UMR6263, Aix-Marseille University,
Marseille, France

Daich A.

URCOM, EA-3221, CNRS FR-3038, UFR Sci.
Tec., Normandie Univ, Le Havre, France

Djebli N.

Pharmacognosy, Api-Phytotherapy Lab.
Mostaganem University, Algeria

El Hatab M.

Natural products Laboratory, Science faculty, Blida
university, Algeria

Esnault M. A.

INRA, UMR 0118 RENN Vegetal Biotechnology
Lab., Rennes, France

Hadj Mahamed M.

BGCMD laboratory, Science Faculty,
Univ. Ouargla, Algérie

Gharabli S.

Chem. Lab., School of App. Med.Sciences,
German Jordanian University, Jordan

Jesus Aizpurua M.

Dept. Organic Chemistry-I, Univ. Basque Country
UPV/EHU, San Sebastian, Spain

Kacimi S.

Materials laboratory, Chemistry dept. Ain
Temouchent University, Algeria

Kessat A.

Analytical Laboratory, Central pharmacy
Rabat, Morocco

Leghseir B.

Phytochemistry laboratory, Faculty of science,
Annaba University, Algeria

Melhaoui A.

LCOMPN-URAC25, Fac. Scie., Mohamed I
University, Oujda, Morocco

Ouahrani M. R.

Faculty of Sciences & Technology, El-Oued
University, Oued Souf, Algeria

Reddy K.H.

Dept. Adv. Res. Center, Narayana Med.College,
Nellore, Andhra Pradesh, India

Salgueiro L.D

Lab. Farmacognosia, Fac. Farmacia, Univ. de
Coimbra, Coimbra, Portugal

Tabcheh M.

Applied Chem. Lab., Faculty of Science Lebanese
University, Tripoli, Lebanon

Villemin D.

LCMT lab., UMR CNRS 6507, ENSICAEN,
Caen, France.

Zyoud A.H.

Dept Chemistry, An-Najah N. University, Nablus,
West Bank, Palestine

Effect of hydrous and saline stress on seeds of *Chamaerops humilis* L. at the stage of germination

Nadjat MÉDJATI^{1,4}, Okkacha HASNAOUI^{2,4*}, Nouria HACHEMI³ & Brahim BABALI^{3,4}

¹ Department of biology, Ahmed Ben Bella University, Oran (Es-Sénia) 31000, Algeria.

² Department of Biology, Dr Tahar Moulay University, Saïda 20000, Algeria

³ Department of Ecology and Environment, Abou Bakr Belkaïd University, Tlemcen 13000, Algeria

⁴ Ecological and Natural Ecosystem Managements Laboratory, Abou Bakr Belkaïd University, Tlemcen 13000, Algeria

Received: November 10, 2016; Accepted: December 30, 2016

Corresponding author Email: okhasnaoui2001@yahoo.fr

Copyright © 2016-POSL

DOI:10.163.pcbjsj/2016.10.3.121

Abstract. In order to later consider a conservation and rehabilitation program, as necessary to evaluate the effect of the main environmental constraints affecting the germination of seeds of *Chamaerops humilis* L., including drought and salinity. For this purpose, we subjected seeds to various concentrations of NaCl (0 to 200 mM) and of PEG 6000 (0 to 200 g/l). The Germination is evaluated by the cumulated seed rate germinated during one 60 day period. The analysis of ANOVA highlighted an effect treatment (concentration of PEG and NaCl) highly significant on the rate of germination ($p \leq 0.05$). The response of seeds of *Chamaerops humilis* to the hydrous and saline stress varies in time with the concentration in PEG and NaCl although it reduces the rate and the speed of germination expressed compared to the witness. Nevertheless, this study shows as well as the germinative capacity of *Chamaerops humilis* with respect to the abiotic constraints is undoubtedly sufficient to retain in the projects of improvement of the pastoral cures and fight against the turning into a desert and to widen consequently its surface of distribution.

Key Words: *Chamaerops humilis* L., Saline stress, Hydrous stress, Germination

1. Introduction

Since more than one decade, the biological consequences of the world climate changes alerted the whole of the scientific community [1-3]. Indeed, a recent scenario of evolution of the world biodiversity indicates that the Mediterranean basin represents one of the eco-regions should undergo the most drastic changes of biodiversity at the dawn of year 2100 [3]. Algeria is part of the group of the Mediterranean countries. These two natural constraints, the drought and salinity, modified the stability of the ecosystems [4] and are mainly the causes of the desertification of the grounds [5]. Under these conditions, the physiology of the plants is disturbed [6-7] and are in large part the causes of desertification of soils [5]; certain spontaneous species disappeared; others are threatened of disappearance [8] and of fall of the outputs [9-10]. With the gleam of these

pessimistic indications, it appears legitimate to wonder about the biological and ecological impacts induced by these stressing environmental conditions on the one hand, and to understand the mechanisms concerned by the plants to adapt to these new environmental conditions. To contribute to the rehabilitation of the affected regions by the aridity and for better fighting against this phenomenon, it is necessary to seek the solutions which make it possible to improve vegetable cover and to solve the problems of regeneration of certain vegetable gasoline's in arid regions, particularly *Chamaerops humilis* (*C. h.*) which is threatened in certain parts of the area of Tlemcen (Western Algeria) [11]. This species held our attention, because it can develop on all soil types, having a system root very developed fixing the road bases of the ground and can be used like means of fight against the erosion of the grounds and the desertification. Compared to the high pharmaceutical value ethno of this taxon [12-15], it is important to control the environmental conditions of its germination to evaluate and limit.

This work aims at the study of the effect of the principal environmental constraints affecting the germination of the seeds [16-17], in particular of the drought and salinity [18-19] which blocks the tests of restoration of the threatened plant species. In this context, we are leaning on the study of germination in conditions of saline and hydrous stress in order to understand the variability of the germination of seeds of *Chamaerops humilis*.

2. Material and Methods

2.1. Plant material

The seeds of *C. h.* were collected on feet of Chamaeropaies, located in the zone of Beni Snous (mounts of Tlemcen), area with Mediterranean climate. The seeds used are selected carefully after a morphological sorting and by using the test of floating to determine viable seeds and their physiological maturity.

2.2 Experimental protocol

2.2.1. Effect of salinity on germination

The seeds selected are peeled their teguments, sterilized during a few minutes in a solution at 1% of hypochlorite of sodium and rinsed additional amount with distilled water. They are then sown in Petri dish 9 cm in diameter and 1.3 cm thickness and are deposited in a regulated plant laboratory with 25°C. Each experimental test relates to 100 seeds at a rate of 5 repetitions of 20 seeds per Petri dish. In each box of kneaded 10 ml of distilled water is versed for pilot seeds and 10 ml of saline solution (NaCl) to various concentrations: 5, 25, 50, 100, and 200 mM for stressed seeds.

The duration of the test was fixed at the period of germination which was spread out over 60 days; the counting of germinated seeds is made daily.

2.2.2. Effect of the hydrous stress on germination

The experimental device of germination of seeds of *C. h.* is the same one as the precedent. The tests of germination were carried out under various levels of hydrous potential simulated using a Polyethylene solution glycol (PEG₆₀₀₀). The PEG constitutes agent relatively stable, inert, nonionic, but quite water soluble. It is not poison even with strong concentrations and the optimal temperature of the germination 25°C. The PEG maintains a potential hydrous stable and uniform during all the experimental period. Indeed, the molecules of PEG₆₀₀₀ constitute a more effective means to simulate a hydrous constraint. The choice of this osmotic agent is justified by its advantages that is to say a product inert, neutral, not affecting the pH and having a raised molecular weight. It does not penetrate in seeds and does not seem to have interferences nor side-effects [20]. Solutions of PEG₆₀₀₀ with increasing concentrations were used to induce the various levels of osmotic stresses tested. The values of the hydrous potential tested are 0, 5, 25, 50, 100 and 200 g/l.

2.3. Statistical analyzes

The results are subjected to the analysis of the variance with only one factor, and the averages are compared by the test of Tukey with the threshold of 5% using the software Minitab16.

3. Results

3.1. Germination in condition of saline stress

3.1.1. Influence saline stress on the rate of germination

The examination of figure 1 illustrating the evolution of the rates of germination according to the increasing NaCl concentrations, watch which the increase in the saline stress actuates a reduction in the germinative capacity expressed compared to the witness. The concentration 5 mM allows a light improvement of germinative capacity which is of 72% compared to the witness (66%). The analysis of ANOVA shows a significant effect of NaCl on the germination of *C. h.* ($p \leq 0.05$).

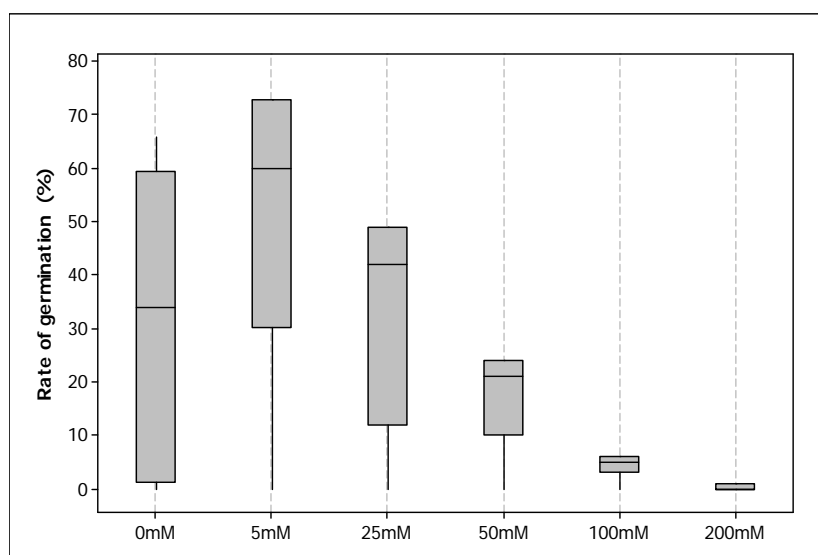


Figure 1. Germination rate of *Chamaerops humilis* under the effect of the saline treatment

3.1.2. Influence saline stress on the kinetics of germination

The kinetics of the germination of seeds under the effect of the increasing NaCl concentrations (Figure 2) described a form sigmoïdale including three phases. The analysis of this kinetics generally shows a first phase of latency, due to the imbibition of seeds, a second exponential phase where one witnesses an acceleration of germination. At the witness, the phase of latency lasts 14 days; the exponential phase of germination lasts 53 days, before reaching the stationary phase where germination stops after a maximum of germination. As the rate of salinity reaches 25, 50, 100 and 200 mM, the shape of this curve is modified in the direction of a stretching, resulting in a delay and a deceleration the speed of germination. However for 5 mM of NaCl, the latency time and the exponential phase are improved respectively to 10 and 44 days.

3.2. Germination in hydrous condition of stress

3.2.1. Influence hydrous stress on the rate of germination

The results concerning the germinative behavior of *C. h.* in terms of germinative capacity under the effect of the various concentrations of PEG appear on figure 2. The figure 3 shows that the germinative capacity of seeds of *Chamaerops* is affected considerably by the hydrous stress. Indeed, more the concentration of the medium in PEG increases more the germinative power

decreases. The rates of germination obtained vary between 66% for the witness and 1% for the medium more concentrated in PEG (200 g/l). Thus the analysis of variance (ANOVA) shows a highly significant effect of hydrous stress on germination ($p \leq 0.05$).

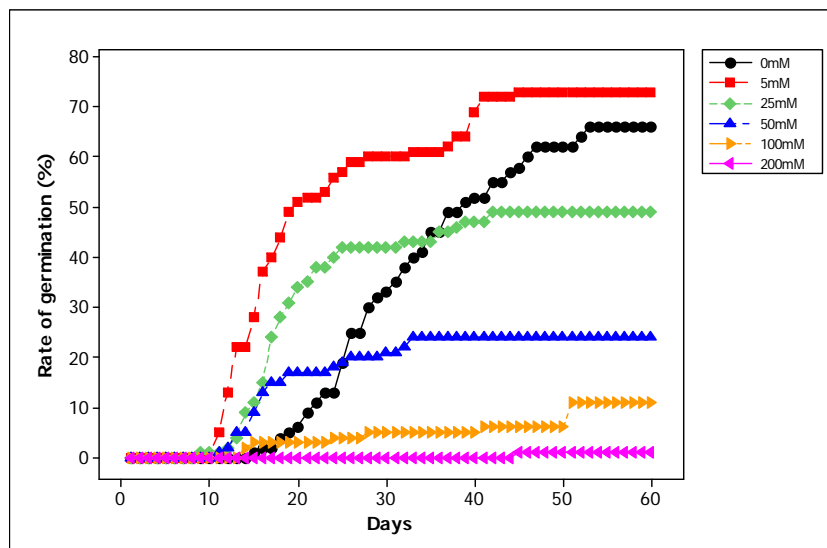


Figure 2. Germination kinetics of *Chamaerops humilis* under the effect of the saline treatment

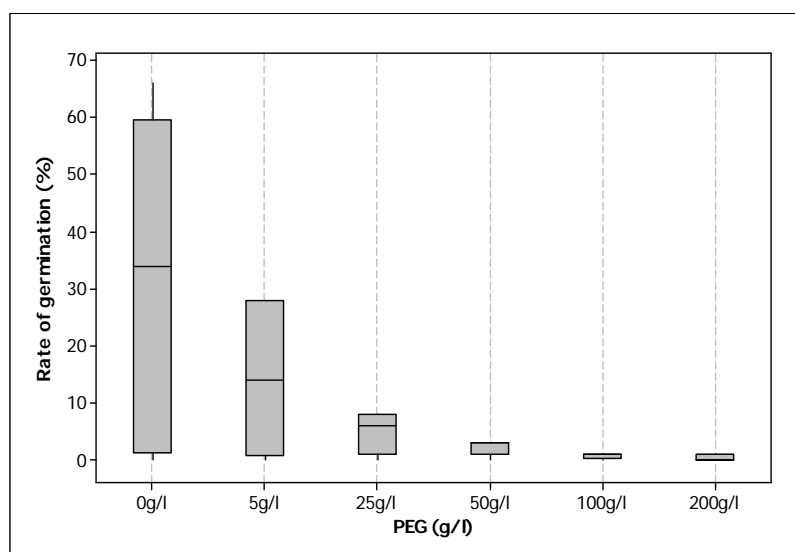


Figure 3. Germination rate of *Chamaerops humilis* under the effect of the osmotic treatment

3.2.2. Influence hydrous stress on the kinetics of germination

The kinetics of germination in conditions of osmotic stress is presented by figure 4, it reflects the sensitivity of the species to the hydrous stress on pilot medium, the curves of kinetics of germination post three phases: latency, acceleration exponentially and finally stage corresponding to a stop of germination after having reached the maximum germinative capacity. The depressive effect of the hydrous deficit on the germination of *C. h.* appears during one or of the whole of these three phases, according to the degree of lowering of the hydrous potential. The effect results in a deceleration the speed of germination visible as of the 5g/l treatment and which is accentuated thereafter.

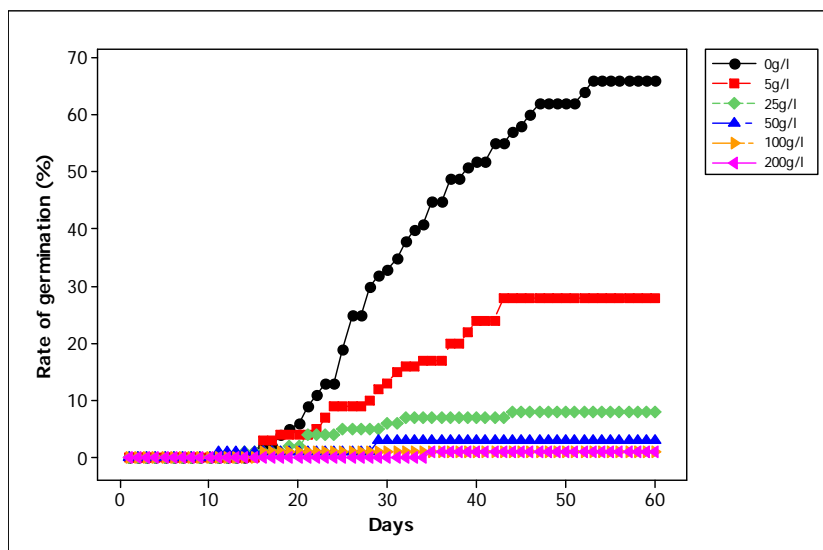


Figure 4. Germination kinetics of *Chamaerops humilis* under the effect of the osmotic treatment

4. Discussion

Studied *Chamaerops humilis* seeds have diverse behaviors towards the osmotic and salt treatment applied during the first phase of development. The results show significant differences in sensitivities of seeds examined against abiotic stresses. Concerning the behavior of the seeds of *C. h.* with regard to salinity, our study shows that NaCl slows down the speed of germination of *C. h.* and decreases their germinative capacity. These effects are all the more marked this salt concentration is high. The germination delay caused by increasing concentrations of the NaCl medium may result from a fix of hydration of seeds in consequence of a high osmotic potential and can be explained by the time necessary with seed to set up mechanisms allowing him to adjust its osmotic pressure interns [21-22] One can notice a relation between the tolerance with salinity at the time of germination and the ecology of each species. In this direction, Neffati [23] announces that the knowledge of the tolerance of salinity at the time of germination is useful information, but nonsufficient to explain the distribution of the species and their development in the salted mediums. The results relating to the effect of the hydrous stress on germination show that the seeds of *C.h.* are fairly tolerant with the hydrous stress. The limiting value of the potential for which the near total of seeds does not germinate any more locates at 100g/l.

The more the osmotic pressure is raised, the more the rate of germination decreases. This study made it possible to note that this species is demanding out of water in germinative phase, but this necessarily does not mean that the tolerant species with the hydrous stress during germination are those which are adapted to the drought at the adult stage [24].

Indeed, although it represents one of the significant factors in the establishment of the species [25-26], the tolerance with the hydrous stress at the time of germination constitutes, according to the conditions which follow this first phase of the vegetative cycle, either an advantage or a disadvantage [27]. The search listings relating to the effect of the hydrous stress on germination show that it is difficult to connect the tolerance to the hydrous constraints, at the time of germination, the ecology of the species even [28]. This result, also noted by Grouzis [27] for certain Sahelian species, makes it possible to say that resistance to the hydrous deficit in germinative phase is not the criterion prevailing of the ecological distribution of tax. Ndour *et al.* [29] affirm that the aptitude to be germinated in conditions of hydrous or saline stress is not obligatorily representative of the ecology of the adult plant, thus joining Sharma [30].

5. Conclusion

The study of the effect of the saline and hydrous stress by the means of NaCl and PEG₆₀₀₀ on the germination of seeds *C. h.* highlighted an effect treatment (concentration of PEG and NaCl) highly significant on the rate of germination ($p \leq 0.05$). Germination in condition of saline stress reveals a good tolerance of the species to salinity since it succeeds in germinating in high NaCl concentrations (to 100 mM, the rate of germination is equal to 11%). In addition, the germinative behavior of the species in hydrous condition of stress shows that this species does not support much the drought at the stage of germination (accentuated inhibition of germination starting from 25g/l). However, it remains necessary to continue these studies at all the stages of development before coming to a conclusion about its tolerance with respect to the saline and hydrous stress. It finally tries to determine if the answer to the pressures applied at the stage of germination constitutes a reliable early indicator of the behavior of the adult plant.

References

- 1- Chapin, F. S., Zavaleta E. S., Eviner V. T. & al. -(2000) Consequences of changing biodiversity. *Nature* 405: 234-242. doi:10.1038/35012241
- 2- Hughes, L. (2000). - Biological consequences of global warming: is the signal already apparent? *Trends Ecol. Evol.*, 15: 56-61.
- 3- Sala, O. E., Chapin, III FS., Armesto, J. J., & al., (2000) . - Global biodiversity scenarios for the year 2100. *Science* 287: 1770-1774.
- 4- Lieth, H., Moshenko, M. & Menzel U. (1997). - Sustainable halophyte utilization in the Mediterranean and Subtropical Dry Regions. *International Conferences on Water management Salinity and Pollution control towards Sustainable Irrigation in the Mediterranean Region*, Valenzano Bari, 23-26 September, 209 p.
- 5- Hamdy, A. (1999). - Saline irrigation and management for a sustainable use. *Advanced Short Course on Saline Irrigation Proceedings*, Agadir (Morocco): 152-227.
- 6- Cramer, GR., Epstein E. & Lauchli A. (1988). - Kinetic of root elongation of maize in response to short term exposure to NaCl and elevated Ca concentration. *J Exp Bot*; 39: 1513-22.
- 7- Belkhodja, M. (1996)- Action de la salinité sur le comportement physiologique, biochimique, hormonal et recherche de marqueurs moléculaires chez la fève (*Vicia faba* L.) *Thèse. Doct. Sci.* Univ. Oran (Algérie), 255 p.
- 8- Gupta, R.K & Abrol, I.P. (1990). - Salts affected soils: their reclamation and management for crop production. *Adv in Soil Science*, 223-88.
- 9- Yeo, A. R., Izard, P., Boursier, P. J., & Flowers, T. J. (1991). Short-and long-term effects of salinity on leaf growth in rice (*Oryza sativa* L.). *Journal of Experimental Botany*, 42(7), 881-889.
- 10- Chevery, C. & Robert, M. (1993). - Salure des sols maghrébins. Influence sur les propriétés physico-chimiques des sols. Répercussions des modifications de ces dernières sur la fertilité, notamment azotée des sols. *Rennes : ENSA*; 59 p.
- 11- Hasnaoui, O., Bouazza, M. & Thinon M. (2006). Contribution à l'étude de la régénération naturelle de *Chamaerops humilis* L.var.*argentea* André dans les zones arides et semi-arides de la région de Tlemcen (Algérie occidentale).*Bull. Soc. Linn. Provence*, T.57
- 12- Beghalia, M., Ghalem, S., Allali, H., Belouatek, A. & Marouf A. (2008). Inhibition of calcium oxalate monohydrate crystal growth using Algerian medicinal plants. *Journal of Medicinal Plants Research*, 2: 66-70.
- 13- Hasnaoui, O., Bouazza, M., Benali, O. & Thinon M. (2011). Ethno botanic study of *Chamaerops humilis* L. Var. *argentea* André (Arecaceae) in western of Algeria. *J. Agricol.* 6(I):1-6.
- 14- Benmehdi, H., Hasnaoui, O., Benali, O. & Salhi F. (2012). Phytochemical investigation of leaves and fruits extracts of *Chamaerops humilis* L. *J. Mater. Environ. Sci.* 3 (2) 320-237.
- 15- Medjati, N. (2014). Contribution à l'étude biologique et phytoécologique du *Chamaerops humilis* L., dans la région occidental de l'Algérie. *Thèse de doctorat : Université de Tlemcen (Algérie)*.
- 16- Come, D. (1970). Les obstacles à la germination. Paris: Masson & Cie.

- 17- Ungar I.A. (1995). Seed germination and seed-bank ecology of halophytes. *In: Kigel J. & Galili G.*, eds. Seed development and germination. New York, USA: Marcel & Dekker Inc
- 18- Ennabli, N. (1995). L'irrigation en Tunisie. *Tunis: Inatdgreg*, 278-30
- 19- Hachicha, M. (2007). - Les sols salés et leur mise en valeur en Tunisie. *Sècheresse*, 18, 45-50.
- 20- Berkat, O. & Briske, D. D. (1982). Water potential evolution of three germination substrates utilizing Polyethylene glycol. *Agronomy journal*, 74, 518-52
- 21- Ben Miled, D., Boussaid, M. & Abdelkefi, A. (1986). Tolérance au sel d'espèces annuelles du genre *Medicago* au cours de germination. *In: Colloque sur les végétaux en milieu aride*, 8-10 septembre 1986, Djerba, Tunisie.
- 22- Smaoui, A. & Cherif, A. (1986). Effet de la salinité sur la germination des graines de cotonnier. *In: Colloque sur les végétaux en milieux arides, 8-10 septembre 1986, Djerba, Tunisie.*
- 23- Nefati, M. (1994). Caractérisation morpho-biologique de certaines espèces végétales nord africaines: implication pour l'amélioration pastorale. *Thèse de doctorat* : Université de Gand (Belgique)
- 24- MC Ginnies, W.J. (1960). Effects of moisture stress and temperature on germination of six range grasses. *Agron. J.*, 52, 159-162.
- 25- MC William, J.R., Clements, R.J. & Dowling, P.M. (1970). - Some factors influencing the germination and early seedling development of pasture plants. *Aust. J. Agric. Res.*, 21, 19-32.
- 26- Boydston, R.A. (1989) - Germination and emergence of longspine sandbur (*Cenchrus longispinus*). *Weed Sci.*, 37, 63-67.
- 27- Grouzis, M. (1987). - Structure, productivité et dynamique des systèmes écologiques sahéliers (mare d'Oursi, Burkina Faso). *Thèse Doct. Sciences Nat*, Université de Paris- Orsay, France, 336 p.
- 28- LE Floc'h, E., Schoenenberegger, A., Nabli, M.A. & Valdeyron G. (1989). Biologie et écologie des principaux taxons. *In: Nabli M.A., Ed. Essai de synthèse sur la végétation et la phyto-écologie tunisienne: I. Éléments de botanique et de phytoécologie*. Tunis : Faculté des Sciences, 51-193.
- 29- Ndour, P. & Danthu, P. (1998). Effet des contraintes hydriques et salines sur la germination de quelques acacias africains. *In : Campa C., Grignon C., Gueye M. & Hamon S., Eds. Colloques et séminaires : l'acacia au Sénégal*. Paris : Orstom, 105-122.
- 30- Sharma, M.L. (1973). Simulation of drought and its effect on germination of five pasture species. *Agron. J.*, 65, 982- 987.

PhytoChem & BioSub Journal

Peer-reviewed research journal on Phytochemistry & Bioactives Substances

ISSN 2170 - 1768



*PCBS
Journal*


ISSN 2170-1768



Edition LPSO - Phytochemistry & Organic Synthesis Laboratory-

<http://www.pcbsj.webs.com>

<https://sites.google.com/site/phytochembsj/>

Email: phytochem07@yahoo.fr